Annals of Research in Antioxidants

Relationship between free radicals and risk of kidney diseases; the role of antioxidants and their reaction mechanisms

Seyed Seifollah Beladi-Mousavi1 , Khadije Hajibabaei2 , Mohammad-Reza Tamadon3 , Mahmoud Rafieian-Kopaei4*

 Chronic Renal Failure Research Center, Ahvaz Junishapur University of Medical Sciences, Ahvaz, Iran Department of Chemistry, Faculty of Sciences, Najafabad Branch, Islamic Azad University, Najafabad, Iran Department of Nephrology, Semnan University of Medical Sciences, Semnan, Iran Medical Plants Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran

Correspondence to:

Prof. Mahmoud Rafieian-Kopaei, Email: Rafieian@yahoo.com

Received: 17 December 2015 **Accepted:** 15 January 2016 **ePublished:** 17 January 2016

Keywords: Free radicals, Reactive oxygen species, Antioxidants, Oxidative stress, Renal Disease

Citation: Beladi-

Mousavi SS, Hajibabaei K, Tamadon MR, Rafieian-Kopaei M. Relationship between free radicals and risk of kidney diseases; the role of antioxidants and their reaction mechanisms. Ann Res Antioxid. 2016;1(1):e02.

Abstract

Free radicals are regularly formed in the human body and are often associated with tissue injury. They are harmful to the body and damage all components of cells, including proteins, DNA, and cell membranes. Antioxidants protect the body from damage caused by free radicals. The lack of balance in production of free radicals and the ability of the body to negate their dangerous effects through neutralization by antioxidants produce oxidative stress (OS). OS has an important role in the pathophysiology of several kidney diseases. There are many experimental evidences suggesting key role of OS and inflammation on renal failure. There are many reports which suggest that the use of antioxidants help in the disease prevention. Therefore, it is very important to understand the reaction mechanism of antioxidant with the free radicals. This review explains the relationship between free radicals and risk of kidney diseases, the role of antioxidants in the diseases prevention and reaction mechanisms of the antioxidants.

Introduction

Free radicals are regularly formed in the human body and often associated with tissue injury. Antioxidants are free radical scavengers that react with the free radicals and delay the cellular damage. The lack of balance in production of free radicals and the ability of the body to negate their dangerous effects through neutralization by antioxidants produce oxidative stress (OS). OS has an important role in the pathophysiology of several kidney diseases. There are many experimental evidences suggesting a key role for OS and inflammation on renal failure ([1,](#page-5-0)[2\)](#page-5-1). There are many reports which suggest that the use of antioxidants help in the disease prevention [\(3](#page-5-2)-[5](#page-5-3)). Hence, it is very important to realize the reaction mechanism of antioxidant with the free radicals. To realize the mechanism of action of antioxidants, it is required to realize the production of free radicals and their harmful reactions. This review explains the generation of free radicals and the damages that free radicals may create and the reaction mechanisms of antioxidants. Also, the relation between OS and risk of kidney diseases and the role of antioxidants in prevention and treatment of these diseases are discussed.

Core tip

Antioxidants protect the body from damage caused by free radicals. The lack of balance in production of free radicals and the ability of the body to negate their dangerous effects through neutralization by antioxidants produce oxidative stress (OS). OS has an important role in the pathophysiology of several kidney diseases. There are many experimental evidences suggesting key role of OS and inflammation on renal failure. There are many reports which suggest that the use of antioxidants help in the disease prevention. Therefore, it is very important to understand the reaction mechanism of antioxidant with the free radicals. This review explains the relationship between free radicals and risk of kidney diseases, the role of antioxidants in the diseases prevention and reaction mechanisms of the antioxidants.

Materials and Methods

For this review, we used a variety of sources by searching through PubMed/Medline, Scopus, EMBASE, EBSCO and directory of open access journals (DOAJ). The search was conducted, using combination of the following key words and, or their equivalents; free radicals, reactive oxygen species,

Copyright © 2016 The Author(s); Published by Society of Diabetic Nephropathy Prevention. This is an open-access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Beladi-Mousavi SS et al

antioxidants, OS and renal disease.

Generation of free radicals in body

Free radicals are molecules with free unpaired electrons. They are very unstable and highly reactive. These compounds are regularly formed in the human body and are often associated with tissue injury. The majority of free radicals are oxygen radicals and other reactive oxygen species (ROS) that are listed in [Table 1](#page-1-0) ([6\)](#page-5-4).

ROS are produced in mitochondrial electron transport system, peroxisomal fatty acids, cytochrome P-450, and phagocytic cells. Drugs, illness, stress, pollution, cigarette smoke, and even exercise can increase free radical exposure. The sources of free radicals formation are showed in [Table 2](#page-1-1) ([7\)](#page-5-5).

Superoxide is produced in mitochondrial electron transport chain by a variety of enzymatic processes such as the NAD(P)H oxidase (Eq. 1).

(1)
$$
2O_2 + NADPH \xrightarrow{(oxidase)}
$$
 $2O_2 + NADP^+ + H^+$

The O_2 is then rapidly converted into H_2O_2 (Eq. 2) by SOD.

(2)
$$
2O_2^{\prime} + 2H^+ \xrightarrow{\text{(SOD)}} H_2O_2 + O_2
$$

'OH is also produced from O_2 ' and H_2O_2 via 'respiratory burst' by Fenton (Eq. 3) and / or Haber-Weiss reactions (Eq. 4) [\(8](#page-5-6)).

 $H_2O_2 + Fe^{+2}$ \longrightarrow 20₂ + H₂O₂ (3)

(4)
$$
2O_2 + H_2O_2
$$
 \longrightarrow OH + OH + O₂

NO is generated from arginine by an enzymatic processes (Eq. 5). The NO• and O_2 ^{\sim} react together to produce peroxynitrite (ONOO) [\(9](#page-5-7)).

- L-Arg + O₂ + NADPH \longrightarrow NO + Citrulline (5)
- $\overrightarrow{NO + O_2}$ \longrightarrow ONOO (6)

Damaging reactions of free radicals in the body

A vast array of molecules are in the human body that more susceptible to free radical attacks than others. These include fats, cellular membranes, DNA, RNA, proteins, carbohydrates and vitamins.

Table 1. Types of the free radicals

Reaction of hydroxyl radical with guanine and the sugar moiety of DNA are shown in [Scheme 1](#page-2-0) and [2](#page-2-1) ([10](#page-5-8)).

Lipid peroxidation is important in vivo and has been widely associated with the tissue injuries and diseases [\(11](#page-5-9)). The mechanism of lipid peroxidation is shown in Eq. 7-10. Generally lipid hydroperoxides are broken down to aldehydes. Commonly aldehydes are biologically active compounds, which attack to the other parts of the cell ([12](#page-5-10)[,13](#page-5-11)).

$$
(7) \quad LH + R' \longrightarrow L' + RH
$$

$$
(8) \quad \dot{L} + O_2 \longrightarrow LOO
$$

$$
(9) \quad \text{LOO}^{\cdot} + \text{LH} \quad \longrightarrow \text{LOOH} + \text{L}^{\cdot}
$$

$$
(10) \quad \text{LOOH} \xrightarrow{\text{LOO + LO + Aldehyde}}
$$

Antioxidants: free radical scavengers

The concept of biological antioxidant relation to any compound that, when existing at a lower concentration compared to that of an oxidizable substrate, is able to either delay or prevent the oxidation of the substrate [\(14\)](#page-5-12). Antioxidant roles include DNA mutations, lowering OS, malignant transformations, as well as other parameters of cell damage.

The body has a defense system for prevention of free radical damage. The first identified types of antioxidant defense systems are those that prevent ROS occurrence and those that block or capture the radicals that are formed. Another antioxidant system of the cell is represented by repair processes which remove the damaged biomolecules before their aggregation enables alteration of cell metabolism [\(15\)](#page-5-13). The repair system intervention consists removing oxidized proteins by proteolytic systems, repairing oxidatively damaged nucleic acids by specific enzymes and repairing oxidized lipids using phospholipases, peroxidases or acyltransferases [\(16](#page-5-14)).

Antioxidants include the enzymatic antioxidants and nonenzymatic antioxidants. These two types of antioxidants modulate the free radical reactions. Body protects itself from free radicals via enzymatic antioxidant mechanisms [\(17\)](#page-5-15). The antioxidant enzymes decrease the levels of lipid hydroperoxide and H_2O_2 , therefore they are important in the prevention of lipid peroxidation and keeping the structure and function of cell membranes. Examples of the enzymatic antioxidants are CAT, GSHPx, SOD, and peroxiredoxin [\(Table 3](#page-2-2)) ([18](#page-5-16),[19](#page-5-17)).

The non-enzymatic antioxidants are of two kinds, the natural antioxidants and the synthetic antioxidants that include vitamins (vitamin E, C, A), bioflavonoids, carotenoids, hydroxycinnamates, theaflavin, theaflavin-3-gallate, allicin, piperine and curcumin. In [Schemes 3](#page-3-0) and [4](#page-3-1) the mechanism of radical scavenging activity of vitamin A, allicin and 2-propenesulfenic acid are shown.

Synthetic antioxidants are added to foods that are sensitive to oxidation to prevent oxidative rancidity. Oxidative rancidity results in demolition of vitamins and essential fatty acids, degradation of flavor, and creation of free radicals that cause stress and damage to our bodies. Synthetic antioxidants which have phenolic structure include butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT) and gallic acid esters [\(23\)](#page-5-18). The antioxidant mechanisms of BHT are shown in [Scheme 5](#page-3-2).

Oxidative stress and diseases

OS is defined as unbalance between the production of free radicals and the ability of the body to counteract their dangerous effects through neutralization by antioxidants [\(24\)](#page-5-19). In certain pathological conditions, increased production of ROS and depletion of antioxidants in defense system leads to enhanced ROS activity and OS, resulting tissue

Scheme 1. Reaction of hydroxyl radical with guanine. **Scheme 2.** Reaction of hydroxyl radical with the sugar moiety of DNA.

damage. OS causes tissue damage by different mechanisms including production of lipid peroxidation, DNA damage, and protein modification. These processes have been related with the pathogenesis of several systemic diseases such as hypertension, diabetes mellitus, and hypercholesterolemia and also kidney disease. In recent years, OS has become one of the most beloved topics in research of molecular mechanism of renal diseases.

Oxidative stress and kidney damage

The kidney is a body part highly vulnerable to damage caused by ROS, likely due to the plenty of long chain polyunsaturated fatty acids in the structure of renal lipids. In recent years, OS has been converted as one of the most popular subjects in research of molecular mechanism of renal disease.

Factors that induce OS in kidney include systemic diseases such as hypertension, diabetes mellitus, hypercholesterolemia, infection, chemotherapeutics, radiocontrast agents environmental toxins, radiation, antibiotics, smoking, occupational chemicals, as well as alcohol consumption. In continuing, we will discuss the relationship between these factors and OS in kidney.

Diabetes mellitus and its association with oxidative stress in kidney

Recent studies have shown that free radicals and kidney damage are associated with diabetic nephropathy. According to this suggestion, diabetes leads to increased glomerular hyperfiltration and a resultant increased glomerular pressure. These lead to damage to glomerular cells and to development of focal and segmental glomerulosclerosis ([25](#page-5-20),[26](#page-5-21)). Angiotensin II inhibitors decrease glomerular pressure and inhibit albuminuria. Increased angiotensin II

Table 3. Enzymatic antioxidants, their cellular locations and the reactions they carry-out

Enzymatic antioxidant	Cellular location	Substrate	Action
GSH	Cytosol	H ₂ O ₂	$H_2O_3 + GSH \rightarrow GSSG + H_2O$
CAT	Peroxisomes cytosol	H _n O _n	$2H_2O_2 \rightarrow O_2 + H_2O$
$PrX-I$	Cytosol	H ₂ O ₂	$H_2O_2 + TrxS_2 \rightarrow Trx(Sh)_2 + H_2O$
Mn/Cu/Zn SOD	Mitochondrial matrix (Mn SOD) cytosol (Cu/Zn SOD)	$O^{\bullet-}$	O^{\bullet} \rightarrow H ₂ O ₂

Scheme 3. Mechanism of radical scavenging activity of vitamin A ([20](#page-5-26)).

Scheme 4. Mechanism for the radical-trapping activity of a) allicin ([21\)](#page-5-27) and b) 2-propenesulfenic acid ([22](#page-5-28)).

Scheme 5. The antioxidant mechanisms of BHT.

level induces OS through activation of NADPH oxidase, stimulating inflammatory cytokines, and so forth [\(27](#page-5-22)[,28\)](#page-5-23). Recently, researches have shown that hyperglycemia-induced OS has role in the pathophysiology of diabetic nephropathy. Lipoic acid as antioxidant improves albuminuria and pathology in diabetes by decrease of OS. Lipoic acid has antioxidant potential and the ability of inhibiting lipid peroxidation [\(29\)](#page-5-24).

Increased blood glucose raises glycosylation of circulator and cellular protein and may incept a series of autoxidation reactions that culminate in the production and accumulation of advanced glycosylation end-products (AGEs) in tissues. The AGEs have oxidizing potential and raise tissue damage by oxygen-free radicals ([30](#page-5-25)).

Hypertension, hypercholesterolemia, fatness, and aging associated with oxidative stress in kidney

Hypertension is one of the main reasons of development of renal failure. Basic cause of this pathology is OS. The most common reason of secondary hypertension is renal artery stenosis that may lead to decrease of renal function and ischemic nephropathy. There is a relation between hypoperfusion and atherosclerosis to interactively increased OS, inflammation and tubular injury in the stenotic kidney ([31](#page-5-29)).

High-fat diet-induced obesity leads to increased hepatic, cardiac and renal tissue OS, which is accompanied by reduction in the antioxidant enzymes activities and glutathione levels that have relation with the increase in MDA and protein carbonyl (PCO) levels ([32](#page-5-30)).

Monocyte chemoattractant protein-1 (MCP-1) is a potent stimulator of macrophage recruitment. It is increased in adipose tissue in obesity and in diabetic kidneys, suggesting that inflammation of these tissues may be MCP-1-dependent ([33](#page-5-31)). Thus we can conclude from these results that macrophages are the reason of increased OS and renal injury in diabetes and obesity-induced renal injury.

Aging is linked with increased OS. Aging cause changes in the kidney such as excessive fibrosis. An increase in apoptosis in cells that control healthy renal functions are often related to excess OS [\(34\)](#page-5-32). At a molecular level, with old increased mutations in nuclear and mitochondrial DNA (mtDNA), increased lipofuscin and AGEs, increased OS, and increased apoptosis have been identified. Proximal tubular cells have large numbers of mitochondria and are the most reliant upon oxidative phosphorylation and most active to oxidant-induced apoptosis and mutations [\(35\)](#page-5-33). Recent researches have shown that anti-aging gen, klotho, has role in renal aging and OS-induced renal damage.

Urinary obstruction, urolithiasis, infection, ischemia reperfusion injury, transplantation of kidney, are associated with oxidative stress in kidney

Most experimental and clinical researches have shown that OS is increased in kidney and systemic circulation. It has been reported that the actions of catalase and manganese superoxide dismutase were raised in early stage of ethylene glycol-induced urolithiasis model in rats. In this experiment, the probable mechanism that leads to free radical raise in the kidney may be different in the course of ethylene glycol-induced urolithiasis. Primarily systemic circulation may bring the toxic substances to the kidney, and finally these substances lead to generate free radicals. In the late step, progressive accumulation of leukocytes and imperfect antioxidant enzyme activities may cause kidney to remain under huge amount of OS ([36,](#page-5-34)[37](#page-5-35)). In experimental urolithiasis studies have shown that decreased antioxidant enzyme activities and involvement of NFκB and p38-MAPK (mitogen-activated protein kinase) signaling pathways are related to OS in rat kidney [\(38](#page-5-36)[-41\)](#page-5-37).

Urinary obstruction and ureteral obstruction owing to urolithiasis is a general urological difficulty seen in urology practice. Unilateral ureteral obstruction (UUO) causes decreased renal CAT and MnSOD protein. UUO-induced nephrotoxicity and renal fibrosis lead to increased OS in kidney.

Infection is another agent that induces OS in kidney. There are many experiments showing that increased OS and re-

duced antioxidant defense mechanisms and antioxidant enzyme systems in kidney may be due to infection [\(42](#page-5-38)[-45\)](#page-5-39). ROS are important mediators that have damaging effects on various organs including kidney during ischemia reperfusion (IR) injury. OS also has a role as a mediator of injury in chronic allograft tubular atrophy and interstitial fibrosis in rat kidney ([46](#page-6-0)).

Renal transplantation is associated with increased OS in kidney in human and animals. Pre-transplant and post-transplant conditions lead to OS increases in transplanted kidney. If there is preexisting diseases such as inflammation, chronic kidney failure, and diabetes mellitus, kidneys are more sensitive to OS during reperfusion injury. Postoperative immunosuppressive agents are among many risk factors of increased OS in kidney [\(47\)](#page-6-1).

Antineoplastic agents, antibiotics, immunosuppressant drugs, analgesics, nonsteroidal anti-inflammatory drugs, and radiocontrast agents are associated with oxidative stress in kidney

Antineoplastic agents are used for the treatment of metastatic cancers. Excess ROS production and depressed antioxidant defense mechanism are responsible for nephrotoxicity. Cisplatin is the well-known and used antineoplastic and nephrotoxic agent. Other nephrotoxic anticancer agents are carboplatin, methotrexate, doxorubicin, cyclosporine and adriamycin.

Antibiotics are nephrotoxic agents that cause induction of OS and depletion of antioxidant enzyme activities in kidney. Researchers showed the protective effects of antioxidants and reactive oxygen scavenger agents against gentamicin-induced nephrotoxicity [\(48](#page-6-2)-[50](#page-6-3)).

Immunosuppressants such as sirolimus and cyclosporine lead to nephrotoxicity via OS ([51](#page-6-4)-[60](#page-6-5)).

Analgesics, particularly paracetamol and nonsteroidal anti-inflammatory drugs (NSAIDs) are extensively used throughout the world. Numerous in vitro and in vivo studies showed that analgesics nephrotoxicity is lead to increased ROS in kidney ([58](#page-6-6)[-60\)](#page-6-5).

Contrast-induced nephropathy (CIN) that is used in imaging procedures is a main clinical concern. CIN is the third main common cause of acute kidney injury in hospitalized patients [\(61](#page-6-7)). Experimental in vitro and vivo researches illustrate raised hypoxia and the formation of ROS inside the kidney following the administration of iodinate contrast media. The use of N-acetyl cysteine and bicarbonate infusion as ROS scavengers lead to reduced ROS in kidney.

Alcohol, smoking, environmental toxins, irradiation and mobile phones associated with oxidative stress in kidney

Ethanol and its metabolites are exorcised into urine, and their amounts in the urine are higher than that of the blood and the liver. Chronic alcohol administration reduces the renal tubular reabsorption and reduces renal function. Functional abnormalities of renal tubules may be related with ethanol-induced changes in membrane composition and lipid peroxidation.

A result of industry is increased air pollution that is another life menacing health problem [\(62\)](#page-6-8). The effect of the pathological changes of diesel exhaust particles (DEPs) on systolic blood pressure (SBP), systemic inflammation, OS, and morphological alterations in lungs, heart, liver, and kidneys in Wistar rats have been shown. So that DEPs lead to inflammation especially in lungs and pulmonary tissue, and these pathological changes are attributed to increased OS and inflammatory cytokines in these tissues ([63](#page-6-9)). Lead and cadmium nephrotoxicity are also associated with increase OS in kidney [\(64,](#page-6-10)[65\)](#page-6-11).

Radiation is an important reason of OS, radiation is commonly used for diagnostic and therapeutic purposes. Chronic OS after total body irradiation leads to radiation nephropathy in rats. The effect of extremely low-frequency electromagnetic field (ELF-EMF) with pulse trains exposure on lipid peroxidation has been shown to lead to OS in the rat liver and kidney tissue. The flow cytometric data proposed a possible association between the exposure to magnetic field and the cell death; however, there were significantly lower necrotic cell percentages in experimental animals compared to either unexposed ([66](#page-6-12)). These results showed the inductive effect of radiation on OS in kidney.

For the last two decades, a large number of studies have investigated the effects of mobile phone radiation on the human and animal. Male reproductive system is among the most affected system ([67,](#page-6-13)[68](#page-6-14)). Increased OS plays a main role in radiofrequency-electromagnetic-waves- (RF-EMW) induced tissue damage.

Conclusion

There is many literatures concerning the association between the OS and renal diseases. Systemic diseases such as diabetes mellitus, hypertension, and hypercholesterolemia; antibiotics, infection; radiocontrast agents and chemotherapeutics; and environmental toxins, radiation, occupational chemicals, smoking, as well as alcohol consumption are associated with induction of OS in the kidney. The kidney is a body part highly vulnerable to damage caused by ROS, due to the plenty of long chain polyunsaturated fatty acids in the structure of renal lipids. Antioxidants have been shown to be effective in animals for protecting kidney. Antioxidant may prevent the OS by peroxidation, inhibiting free radicals and also via other mechanism which can inhibit diseases.

Authors' contribution

Primary draft by SH. SSBM and MRT researching the data and conducted primary editing. Editing the final manuscript by MRK.

Conflicts of interest

The authors declared no competing interests.

Ethical considerations

Ethical issues (including plagiarism, data fabrication, double publication) have been completely observed by the authors.

Funding/Support None.

Beladi-Mousavi SS et al

References

- 1. Himmelfarb J, Hakim RM. Oxidative stress in uremia. Curr Opin Nephrol Hypertens. 2003;12:593-8.
- 2. Galle J. Oxidative stress in chronic renal failure. Nephrol Dial Transplant. 2001;16:2135-7.
- 3. Willcox JK, Ash SL, Catignani GL. Antioxidants and prevention of chronic disease. Crit Rev Food Sci Nutr. 2004;44:275-95.
- 4. Hajhashemi V, Vaseghi G, Pourfarzam M, Abdollahi A. Are antioxidants helpful for disease prevention? Res Pharm Sci. $2010.5.1 - 8$
- 5. Halliwell B. How to characterize an antioxidant: an update. Biochem Soc Symp. 1995;61:73-101.
- 6. Bashan N, Kovsan J, Kachko I, Ovadia H, Rudich A. Positive and negative regulation of insulin signaling by oxygen and nitrogen species. Physiol Rev. 2009;89:27-71.
- 7. Bast A, Haenen GR, Doelman CJ. Oxidants and antioxidants: state of the art. Am J Med. 1991;91:2S-13S.
- 8. Knight JA. In Free Radicals, Antioxidants, Aging and Disease. Washington: AACC Press; 1999.
- 9. Zhu L, Gunn C, Beckman JS. Bactericidal activity of peroxynitrite. Arch Biochem Biophys. 1992;298:452-7.
- 10. Dizdaroglu M, Jaruga P, Birincioglu M, Rodriguez H. Free radical-induced damage to DNA: mechanisms and measurement. Free Rad. Biol Med. 2002;32:1102-15.
- 11. Esterbauer H, Schaur RJ, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. Free Radic Biol Med. 1991;11:81-128.
- 12. Pryor WA, Porter NA. Suggested mechanisms for the production of 4-hydroxy-2-nonenal from the autoxidation of polyunsaturated fatty acids. Free Radic Biol Med. 1990;8:541- 3.
- 13. Devasgayam TP, Boloor KK, Ramasarma T. Methods for estimating lipid peroxidation: an analysis of merits and demerits. Indian J Biochem Biophys. 2003;40: 300-8.
- 14. Nasri H, Hajian S, Ahmadi A, Baradaran A, Kohi G, Nasri P, et al. Ameliorative effect of green tea against contrast-induced renal tubular cell injury. Iran J Kidney Dis. 2015;9:421-6.
- 15. Cheeseman KH, Slater TF. An introduction to free radical biochemistry. Br Med Bull. 1993;49:481-93.
- 16. Hitchon CA, El-Gabalawy HS. Oxidation in rheumatoid arthritis. Arthritis Res Ther. 2004;6:265-78.
- 17. Koruk M, Taysi S, Savas MC, Yilmaz O, Akcay F, Karakok M. Oxidative stress and enzymatic antioxidant status in patients with nonalcoholic steatohepatitis. Ann Clin Lab Sci. 2004;34: 57-62.
- 18. Zhan CD, Sindhu RK, Pang J, Ehdaie A, Vaziri ND. Superoxide dismutase, catalase and glutathione peroxidase in the spontaneously hypertensive rat kidney: effect of antioxidantrich diet. J Hypertens. 2004;22:2025-33.
- 19. Stone JR, Yang S. Hydrogen peroxide: a signaling messenger. Antioxid Redox Signal. 2006;8:243-70.
- 20. Livrea MA, Tesoriere L, Bongiorno A, Pintaudi AM, Ciaccio M, Riccio A. Contribution of vitamin A to the oxidation resistance of human low density lipoproteins. Free Radic Biol Med. 1995;18:401-9.
- 21. Amorati R, Lynett PT, Valgimigli L, Pratt DA. The Reaction of Sulfenic Acids with Peroxyl Radicals: Insights into the Radical-Trapping Antioxidant Activity of Plant-Derived Thiosulfinates. Chem.–A Eur. J. 2012; 18:6370–6379.
- 22. Galano A, Francisco-Marquez M. Peroxyl-radical-scavenging activity of garlic: 2-propenesulfenic acid versus allicin. J Phys Chem B. 2009;113:16077-81.
- 23. Nasri H, Shirzad H, Baradaran A, Rafieian-Kopaei M. Antioxidant plants and diabetes mellitus. J Res Med Sci. 2015;20:491-502.
- 24. Nasri H, Ardalan MR, Rafieian-Kopaei M. Mechanistic Impacts of Medicinal Plants in Diabetic Kidney Disease. Iran J Public Health. 2014;43:1311-3.
- 25. Anderson S, Brenner BM. Pathogenesis of diabetic glomerulopathy: hemodynamic considerations. Diabetes

Metab Rev. 1988;4:163-77.

- 26. Zatz R, Dunn BR, Meyer TW. Prevention of diabetic glomerulopathy by pharmacological amelioration of glomerular capillary hypertension. J Clin Invest. 1986; 77:1925-30.
- 27. Garrido AM, Griendling KK. NADPH oxidases and angiotensin II receptor signaling. Mol Cell Endocrinol. 2009;302:148-58.
- 28. Mehta PK, Griendling KK. Angiotensin II cell signaling: physiological and pathological effects in the cardiovascular system. Am J Physiol. 2007;292:C82-97.
- 29. Dinçer Y, Telci A, Kayalı, Yılmaz LA, Çakatay U, Akçay T. Effect of alpha-lipoic acid on lipid peroxidation and anti-oxidant enzyme activities in diabetic rats. Clin Exp Pharmacol Physiol. 2002;29:281-4.
- 30. Mansouri E, Panahi M, Ghaffari MA, Ghorbani A, Effects of grape seed proanthocyanidin extract on oxidative stress induced by diabetes in rat kidney. Iran Biomed J. 2011;15:100- 6.
- 31. Chade AR, Rodriguez-Porcel M, Grande JP. Dis-tinct renal injury in early atherosclerosis and renovascular disease. Circulation. 2002;106:1165-71.
- 32. Noeman SA, Hamooda HE, Baalash AA. Biochemical study of oxidative stress markers in the liver, kidney and heart of high fat diet induced obesity in rats. Diabetol Metab Syndr. 2011;3:1-17.
- 33. Chow FY, Nikolic-Paterson DJ, Ma FY, Ozols E, Rollins BJ, Tesch GH. Monocyte chemoattractant protein-1-induced tissue inflammation is critical for the development of renal injury but not type 2 diabetes in obese db/db mice. Diabetologia. 2007; 50:471-80.
- 34. Zuk A, Bonventre JV. Acute Kidney Injury. Annu Rev Med. 2016;67:293-307.
- 35. Percy CJ, Power D, Gobe GC. Renal ageing: changes in the cellular mechanism of energy metabolism and oxidant handling. Nephrology. 2008;13:147-52.
- 36. Huang HS, Ma MC, Chen J, Chen CF. Changes in the oxidantantioxidant balance in the kidney of rats with nephrolithiasis induced by ethylene glycol. J Urol. 2002;167:2584-93.
- 37. Huang HS, Chen CF, Chien CT, Chen J. Possible biphasic changes of free radicals in ethylene glycolinduced nephrolithiasis in rats. BJU Int. 2000; 85:1143-9.
- 38. Ilbey YO, Ozbek E, Sims A, Cekmen M, Somay A, Tasci AI. Pyrrolidine dithiocarbamate treatment prevents ethylene glycol-induced urolithiasis through inhibition of NF-kB and p38-MAPK signaling pathways in rat kidney. Arch Ital Urol Androl. 2010;82:87-94.
- 39. Ilbey YO, Ozbek E, Simsek A, Cekmen M, Somay A, Tasci AI. Effects of pomegranate juice on hyperoxaluriainduced oxidative stress in the rat kidneys. Ren Fail. 2009;31:522-31.
- 40. Tugcu V, Kemahli E, Ozbek E. Protective effect of a potent antioxidant, pomegranate juice, in the kidney of rats with nephrolithiasis induced by ethylene glycol. J Endourol. 2008; 22:2723-31.
- 41. TugcuV, Ozbek E, Kemahli E. Rapid communication: protective effect of a nuclear factor kappa B inhibitor, pyrolidium dithiocarbamate, in the kidney of rats with nephrolithiasis induced by ethylene glycol. J Endourol. 2007;21:1097-06.
- 42. Tothova L, Hodosy J, Kamodyova N. Bactofection with toll-like receptor 4 in a murine model of urinary tract infection. Curr Microbiol. 2011; 62:1739–1742.
- 43. Han CH, Kim SH, Kang SH. Protective effects of cranberries on infection-induced oxidative renal damage in a rabbit model of vesico-ureteric reflux. BJU Int. 2007;100:1172-5.
- 44. Celik S, Gorur S, Aslantas O, Erdogan S, Ocak S, Hakverdi S. Caffeic acid phenethyl ester suppresses oxidative stress in Escherichia coli pyelonephritis in rats. Mol Cell Biochem. 2007;297:131-8.
- 45. Sener G, Tuğtepe H, Velioğlu-Oğünç A, Cetinel S, Gedik N, Yeğen BC. Melatonin prevents neutrophil-mediated oxidative injury in Escherichia coli-induced pyelonephritis in rats. J

Pineal Res. 2006;41:220-7.

- 46. Djamali A. Oxidative stress as a common pathway to chronic tubulointerstitial injury in kidney allografts. Am J Physiol. 2007;293:F445–55.
- 47. Nafar M, Sahraei Z, Salamzadeh J, Samavat S, Vaziri ND. Oxidative stress in kidney transplantation: causes, consequences, and potential treatment. Iran J Kidney Dis. 2001;5:357-72.
- 48. Maniu A. Perde-Schrepler M, Cosgarea M. Protective effect of L-N-acetylcysteine against gentamycin ototoxicity in the organ cultures of the rat cochlea. Roman J Morphol Embryol. 2011;52:159-64.
- 49. Ozbek E, Turkoz Y, Sahna E, Ozugurlu F, Mizrak B, Ozbek M. Melatonin administration prevents the nephrotoxicity induced by gentamicin. BJU Int. 2000;85:742-6.
- 50. Ozbek E, Ilbey YO, Simsek A, Cekmen M, Mete F, Somay A. Rosiglitazone, peroxisome proliferator receptorgamma agonist, ameliorates gentamicin-induced nephrotoxicity in rats. Int Urol Nephrol. 2010;42:579-87.
- 51. Giustarini D, Dalle-Donne I, Paccagnini E, Milzani A, Rossi R. Carboplatin-induced alteration of the thiol homeostasis in the isolated perfused rat kidney. Arch Biochem Biophys. 2009;488:83-9.
- 52. Schmitz V, Klawitter J, Bendrick-Peart J. Metabolic profiles in urine reflect nephrotoxicity of sirolimus and cyclosporine following rat kidney transplantation. Nephron Exp Nephrol. 2009;111:e80-91.
- 53. Kolli VK, Abraham P, Isaac B, Selvakumar D. Neutrophil infiltration and oxidative stress may play a critical role in methotrexate-induced renal damage. Chemotherapy. 2009;55:83-90.
- 54. Ajith TA, Aswathy MS, Hema U. Protective effect of Zingiber officinale roscoe against anticancer drug doxorubicin-induced acute nephrotoxicity. Food Chem Toxical. 2008;46:3178-81.
- 55. Galletti P, di Gennaro CI, Migliardi V. Diverse effects of natural antioxidants on cyclosporin cytotoxicity in rat renal tubular cells. Nephrol Dial Transplant. 2005;20:1551-8.
- 56. Malarkodi KP, Balachandar AV, Varalakshmi P. Protective effect of lipoic acid on adriamycin induced lipid peroxidation in rat kidney. Mol Cell Biochem. 2003;247:9-13.
- 57. Ates A, Ahin S, Ceribas AO, Yilmaz S. Lycopene, a carotenoid,

attenuates cyclosporine-induced renal dysfunction and oxidative stress in rats. Basic Clin Pharmacol Toxicol. 2007;100:372-6.

- 58. Tirkey N, Kaur G, Vij G, Chopra K. Curcumin, a diferuloylmethane, attenuates cyclosporine-induced renal dysfunction and oxidative stress in rat kidneys. BMC Pharmacol. 2005;5:15.
- 59. Naghizadeh B, Mansouri SMT, Mashhadian NV. Crocin attenuates cisplatin-induced renal oxidative stress in rats. Food Chem Toxical. 2010;48:2650-5.
- 60. Kalaiselvi P, Pragasam V, Chinnikrishnan S, Veena CK, Sundarapandiyan R, Varalakshmi P. Counteracting adriamycininduced oxidative stress by administration of N-acetyl cysteine and vitamin E. Clin Chem Lab Med. 2005;43:834-40.
- 61. Cronin RE. Contrast-induced nephropathy: pathogenesis and prevention. Pediatr Nephrol. 2010;25:191-204.
- 62. Boor P, Casper S, Celec P. Renal, vascular and cardiac fibrosis in rats exposed to passive smoking and industrial dust fibre amosite. J Cell Mol Med. 2009;13:4484-91.
- 63. Nemmar A, Al-Salam S, Zia S, Dhanasekaran S, Shudadevi M, Ali BH. Time-course effects of systemically administered diesel exhaust particles in rats. Toxicol Lett. 2010;194:58-65.
- 64. Navarro-Moreno LG, Quintanar-Escorza MA, Gonzalez S. Effects of lead intoxication on intercellular junctions and biochemical alterations of the renal proximal tubule cells. Toxicol In Vitro. 2009;23:1298-304.
- 65. Wang L, Cao J, Chen D, Liu X, Lu H, Liu Z. Role of oxidative stress, apoptosis, and intracellular homeostasis in primary cultures of rat proximal tubular cells exposed to cadmium. Biol Trace Elem Res. 2009;127:53-68.
- 66. Lenarczyk M, Cohen EP, Fish BL. Chronic oxidative stress as a mechanism for radiation nephropathy. Radiat Res. 2009; 171:164-72.
- 67. Desai NR, Kesari KK, Agarwal A. Pathophysiology of cell phone radiation: oxidative stress and carcinogenesis with focus on male reproductive system. Reprod Biol Endocrinol. 2009;22:114.
- 68. Agarwal A1, Desai NR, Makker K, Varghese A, Mouradi R, Sabanegh E, et al. Effects of radiofrequency electromagnetic waves (RF-EMW) from cellular phones on human ejaculated semen: an in vitro pilot study. Fertil Steril. 2009;92:1318-25.